10.11609/jott.2023.15.10.23931-24150 www.threatenedtaxa.org

> 26 October 2023 (Online § Print) 15(10): 23931-24150 ISSN 0974-79t07 (Online) ISSN 0974-7893 (Print)



Open Access

# Rourde conservation globally Journal of Threatened Taxa



## ISSN 0974-7907 (Online); ISSN 0974-7893 (Print)

Publisher

Wildlife Information Liaison Development Society www.wild.zooreach.org

Host **Zoo Outreach Organization** www.zooreach.org

43/2 Varadarajulu Nagar, 5th Street West, Ganapathy, Coimbatore, Tamil Nadu 641006, India Registered Office: 3A2 Varadarajulu Nagar, FCI Road, Ganapathy, Coimbatore, Tamil Nadu 641006, India Ph: +91 9385339863 | www.threatenedtaxa.org

Email: sanjay@threatenedtaxa.org

#### EDITORS

Founder & Chief Editor

#### Dr. Sanjay Molur

Wildlife Information Liaison Development (WILD) Society & Zoo Outreach Organization (ZOO), 43/2 Varadarajulu Nagar, 5th Street West, Ganapathy, Coimbatore, Tamil Nadu 641006, India

## **Deputy Chief Editor**

Dr. Neelesh Dahanukar Noida, Uttar Pradesh, India

#### Managing Editor

Mr. B. Ravichandran, WILD/ZOO, Coimbatore, Tamil Nadu 641006, India

#### Associate Editors

Dr. Mandar Paingankar, Government Science College Gadchiroli, Maharashtra 442605, India Dr. Ulrike Streicher, Wildlife Veterinarian, Eugene, Oregon, USA Ms. Privanka Iver. ZOO/WILD. Coimbatore. Tamil Nadu 641006. India Dr. B.A. Daniel, ZOO/WILD, Coimbatore, Tamil Nadu 641006, India

#### **Editorial Board**

Dr. Russel Mittermeier

Executive Vice Chair, Conservation International, Arlington, Virginia 22202, USA

#### Prof. Mewa Singh Ph.D., FASc, FNA, FNASc, FNAPsy

Ramanna Fellow and Life-Long Distinguished Professor, Biopsychology Laboratory, and Institute of Excellence, University of Mysore, Mysuru, Karnataka 570006, India; Honorary Professor, Jawaharlal Nehru Centre for Advanced Scientific Research, Bangalore; and Adjunct Professor, National Institute of Advanced Studies, Bangalore

#### Stephen D. Nash

Scientific Illustrator, Conservation International, Dept. of Anatomical Sciences, Health Sciences Center, T-8, Room 045, Stony Brook University, Stony Brook, NY 11794-8081, USA

**Dr. Fred Pluthero** 

#### Toronto, Canada

Dr. Priya Davidar

Sigur Nature Trust, Chadapatti, Mavinhalla PO, Nilgiris, Tamil Nadu 643223, India

#### Dr. Martin Fisher

Senior Associate Professor, Battcock Centre for Experimental Astrophysics, Cavendish Laboratory, JJ Thomson Avenue, Cambridge CB3 OHE, UK

#### **Dr. John Fellowes**

Honorary Assistant Professor, The Kadoorie Institute, 8/F, T.T. Tsui Building, The University of Hong Kong, Pokfulam Road, Hong Kong

#### Prof. Dr. Mirco Solé

Universidade Estadual de Santa Cruz, Departamento de Ciências Biológicas, Vice-coordenador do Programa de Pós-Graduação em Zoologia, Rodovia Ilhéus/Itabuna, Km 16 (45662-000) Salobrinho. Ilhéus - Bahia - Brasil

#### Dr. Rajeev Raghavan

Professor of Taxonomy, Kerala University of Fisheries & Ocean Studies, Kochi, Kerala, India

## **English Editors**

Mrs. Mira Bhojwani, Pune, India Dr. Fred Pluthero, Toronto, Canada Mr. P. Ilangovan, Chennai, India Ms. Sindhura Stothra Bhashyam, Hyderabad, India

#### Web Development

Mrs. Latha G. Ravikumar, ZOO/WILD, Coimbatore, India

#### Typesetting

Mrs. Radhika, ZOO, Coimbatore, India Mrs. Geetha, ZOO, Coimbatore India

Fundraising/Communications Mrs. Payal B. Molur, Coimbatore, India

#### Subject Editors 2020-2022

#### Fungi

- Dr. B. Shivaraju, Bengaluru, Karnataka, India
- Dr. R.K. Verma, Tropical Forest Research Institute, Jabalpur, India
- Dr. Vatsavaya S. Raju, Kakatiay University, Warangal, Andhra Pradesh, India
- Dr. M. Krishnappa, Jnana Sahyadri, Kuvempu University, Shimoga, Karnataka, India Dr. K.R. Sridhar, Mangalore University, Mangalagangotri, Mangalore, Karnataka, India
- Dr. Gunjan Biswas, Vidyasagar University, Midnapore, West Bengal, India
- Dr. Kiran Ramchandra Ranadive, Annasaheb Magar Mahavidyalaya, Maharashtra, India

#### Plants

- Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India
- Dr. N.P. Balakrishnan, Ret. Joint Director, BSI, Coimbatore, India
- Dr. Shonil Bhagwat, Open University and University of Oxford, UK
- Prof. D.J. Bhat, Retd. Professor, Goa University, Goa, India
- Dr. Ferdinando Boero, Università del Salento, Lecce, Italy
- Dr. Dale R. Calder, Royal Ontaro Museum, Toronto, Ontario, Canada
- Dr. Cleofas Cervancia, Univ. of Philippines Los Baños College Laguna, Philippines
- Dr. F.B. Vincent Florens, University of Mauritius, Mauritius
- Dr. Merlin Franco, Curtin University, Malaysia
- Dr. V. Irudayaraj, St. Xavier's College, Palayamkottai, Tamil Nadu, India
- Dr. B.S. Kholia, Botanical Survey of India, Gangtok, Sikkim, India
- Dr. Pankaj Kumar, Department of Plant and Soil Science, Texas Tech University, Lubbock, Texas, USA.
- Dr. V. Sampath Kumar, Botanical Survey of India, Howrah, West Bengal, India
- Dr. A.J. Solomon Raju, Andhra University, Visakhapatnam, India
- Dr. Vijayasankar Raman, University of Mississippi, USA Dr. B. Ravi Prasad Rao, Sri Krishnadevaraya University, Anantpur, India
- Dr. K. Ravikumar, FRLHT, Bengaluru, Karnataka, India
- Dr. Aparna Watve, Pune, Maharashtra, India
- Dr. Qiang Liu, Xishuangbanna Tropical Botanical Garden, Yunnan, China
- Dr. Noor Azhar Mohamed Shazili, Universiti Malaysia Terengganu, Kuala Terengganu, Malaysia
- Dr. M.K. Vasudeva Rao, Shiv Ranjani Housing Society, Pune, Maharashtra, India
- Prof. A.J. Solomon Raju, Andhra University, Visakhapatnam, India
- Dr. Mandar Datar, Agharkar Research Institute, Pune, Maharashtra, India
- Dr. M.K. Janarthanam, Goa University, Goa, India Dr. K. Karthigeyan, Botanical Survey of India, India
- Dr. Errol Vela, University of Montpellier, Montpellier, France
- Dr. P. Lakshminarasimhan, Botanical Survey of India, Howrah, India Dr. Larry R. Noblick, Montgomery Botanical Center, Miami, USA
- Dr. K. Haridasan, Pallavur, Palakkad District, Kerala, India
- Dr. Analinda Manila-Fajard, University of the Philippines Los Banos, Laguna, Philippines
- Dr. P.A. Sinu, Central University of Kerala, Kasaragod, Kerala, India
- Dr. Afroz Alam, Banasthali Vidyapith (accredited A grade by NAAC), Rajasthan, India
- Dr. K.P. Rajesh, Zamorin's Guruvayurappan College, GA College PO, Kozhikode, Kerala, India
- Dr. David E. Boufford, Harvard University Herbaria, Cambridge, MA 02138-2020, USA
- Dr. Ritesh Kumar Choudhary, Agharkar Research Institute, Pune, Maharashtra, India
- Dr. A.G. Pandurangan, Thiruvananthapuram, Kerala, India

Dr. Navendu Page, Wildlife Institute of India, Chandrabani, Dehradun, Uttarakhand, India Dr. Kannan C.S. Warrier, Institute of Forest Genetics and Tree Breeding, Tamil Nadu, India

#### Invertebrates

- Dr. R.K. Avasthi, Rohtak University, Haryana, India
- Dr. D.B. Bastawade, Maharashtra, India
- Dr. Partha Pratim Bhattacharjee, Tripura University, Suryamaninagar, India
- Dr. Kailash Chandra, Zoological Survey of India, Jabalpur, Madhya Pradesh, India
- Dr. Ansie Dippenaar-Schoeman, University of Pretoria, Queenswood, South Africa Dr. Rory Dow, National Museum of natural History Naturalis, The Netherlands
- Dr. Brian Fisher, California Academy of Sciences, USA
- Dr. Richard Gallon, llandudno, North Wales, LL30 1UP
- Dr. Hemant V. Ghate, Modern College, Pune, India
- Dr. M. Monwar Hossain, Jahangirnagar University, Dhaka, Bangladesh

For Focus, Scope, Aims, and Policies, visit https://threatenedtaxa.org/index.php/JoTT/aims_scope For Article Submission Guidelines, visit https://threatenedtaxa.org/index.php/JoTT/about/submissions For Policies against Scientific Misconduct, visit https://threatenedtaxa.org/index.php/JoTT/policies_various	
,	continued on the back inside cover

\_\_\_\_\_

Journal of Threatened Taxa | www.threatenedtaxa.org | 26 October 2023 | 15(10): 24063-24078 ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print) OPEN ACCESS https://doi.org/10.11609/jott.8420.15.10.24063-24078 (I)  $(\alpha)$ #8420 | Received 23 February 2023 | Final received 05 June 2023 | Finally accepted 27 September 2023 

# A compendium of *Aphelenchoides* (Fischer, 1894) (Nematoda: Tylenchina: Aphelenchoidea) nematodes with the description of a new species from Manipur, India

## Loukrakpam Bina Chanu<sup>1</sup> 🗈 & Naorem Mohilal<sup>2</sup> 🗈

<sup>1,2</sup> Department of Zoology, Manipur University, Canchipur, Manipur 795003, India. <sup>1</sup> bina.chanu@gmail.com (corresponding author), <sup>2</sup> nmohilal@manipuruniv.ac.in

Abstract: The present compendium is based on the findings of a research work on the survey of nematodes belonging to the family Aphelenchoidea in the northeastern states of India and the literature available on this particular species, mainly from Manipur. During the study, a total of 12 Aphelenchoides spp. were found, among which six species were reported for the first time from Manipur. A new species, Aphelenchoides oryzae is also described in the present article. The present study will help in making us understand the biodiversity status of Aphelenchoides nematodes in the region. Diagnosis of the species and illustrations along with dichotomous keys are provided in the manuscript.

Keywords: Aphelenchid, Coconut tree, fungivore, food-web, soil fertility, soil dwelling, Pinus sp., Morus sp., Orange plant and species richness.

ZooBank: urn:lsid:zoobank.org:pub:517493EB-10A5-4707-A288-4557240ED5EB

Editor: Biplob K. Modak, Sidho Kanho Birsha University, West Bengal, India.

Citation: Chanu, L.B. & N. Mohilal (2023). A compendium of Aphelenchoides (Fischer, 1894) (Nematoda: Tylenchina: Aphelenchoidea) nematodes with the description of a new species from Manipur, India. Journal of Threatened Taxa 15(10): 24063–24078. https://doi.org/10.11609/jott.8420.15.10.24063-24078

Copyright: © Chanu & Mohilal 2023. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

Funding: The study was made under the funding agency of SERB, New Delhi with file no. SB/FT/LS-113/2013.

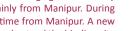
Competing interests: The authors declare no competing interests.

Author details: L. BINA CHANU is a guest faculty at the Department of Life Sciences (Zoology), Manipur University, Canchipur. She is working on biodiversity of nematode parasites including ectoparasites. N. MOHILAL is a professor, Department of Life Sciences (Zoology), Manipur University, Canchipur. He works on the biodiversity of soil nematodes and their bio-control methods using medicinal plants, biodiversity of Annelids, snakes and birds.

Author contributions: LBC conducted the survey, collected samples, processed the samples, extracted nematodes, identified the nematodes, did statistical calculations and wrote the manuscript. NM helped in designing the survey and collection, in the identification of the nematodes, and in final proofing of the manuscript.

Acknowledgements: The authors are thankful to the Head of Department, Department of Zoology, Manipur University, Canchipur, Manipur, India for providing necessary laboratory facilities and the Science & Engineering Board, New Delhi for the funding with the no. SB/FT/LS-113/2013.





Date of publication: 26 October 2023 (online & print)



## INTRODUCTION

Forest conservation improves ecosystem functions and will help to protect natural biodiversity. Plant communities are the critical indicators for forest restoration. Below-ground diversity relates verv closely with above-ground biodiversity. Plant parasitic nematodes are found in every soil of varied ecosystems. Nematodes are often sensitive to habitat disturbance, showing the characteristic sequence of recolonization after disturbance. Furthermore, they are represented in a wide array of trophic groups as herbivores (Tylenchid), bacterivores & fungivores (Aphelenchid), omnivores (Dorylaimid), and predators (Mononchid) reflecting resource availability and changes of environmental conditions in the soil providing information on succession and changes in decomposition pathways in the soil food - web, nutrient status & soil fertility, acidity, and the effects of soil contaminants (Yeates & Bongers 1999).

Bacterial as well as fungal feeding nematodes like aphelenchids have a high carbon: nitrogen (C:N) ratio (±5.9) than their substrate (±4.1), consuming bacteria, they take in more N than necessary for their body structure. The excess nitrogen is excreted as ammonia. The bacterial and fungal feeding nematodes' community in the top 15 cm of the field soil mineralizes N at rates increasing to  $1.01\mu g - N g - soil - 1d - 1$  in the rhizosphere (Ferris et al. 1995; 1996; 1997).

Aphelenchid nematodes have diverse habitats. Several aphelenchids are associated with insects, some spending a part of their life - cycle in insects besides being phytophagous, while others are mycetophagous. Some forms of aphelenchids are true plant parasites and are, therefore, economically significant. Of the available aphelenchid nematodes, three species are major pests of agricultural and horticultural crops, i.e., white-tip nematode *Aphelenchoides besseyi*; Red-ring nematode of coconut *Rhadinaphelenchus cocophilus*, and pine-wilt nematode *Bursaphelenchus xylophilus* respectively in the world. So far, 138 species of *Aphelenchoides* (Fischer, 1894) have been identified, of which India contributed more than 12 species.

Recognizing the importance of bacterial and fungal decomposition in forest ecosystems, survey for Aphelenchid nematodes is very important in every region of the Earth. Recently, a survey was conducted on the *Aphelenchoides* nematodes in the different ecosystems of the north-eastern region of India. During the work, 12 *Aphelenchoides* were encountered from Manipur, among which six species were recorded for the first time from this particular region. The species encountered were Aphelenchoides aligarhiensis (Siddiqi et al., 1967); A. baquei (Maslen, 1978); A. confusus (Thorne & Malek, 1968); A. minor (Seth & Sharma, 1986); A. swarupi (Seth & Sharma, 1986); A. vigor (Thorne & Malek, 1968); A. dhanachnadi (Chanu et al., 2012) and A. neoechinocaudatus (Chanu et al., 2012); A. aerialis (Chanu et al., 2015); A. longistylus (Chanu & Mohilal, 2014); A. neominoris (Chanu & Mohilal, 2014), and A. manipurensis (Chanu & Mohilal, 2018). These species are presented along with their dichotomous keys.

## METHODS

#### Study site

## Methodology

For collection of soil samples around the rhizospheric region of a particular host, 500 g of soil around the plant from 8 different sides were taken. The soil was mixed together thoroughly. From the thoroughly mixed soil, again 500 g were taken, serving as the sample soil for a particular host plant or tree. The samples were processed for extraction of nematodes through the Cobb's (1918) sieving and decanting method and Baermann's funnel technique. Collected nematodes through the process were fixed with warm formalin alcohol (F.A) (4:1) for 24 hours and afterward, dehydrated under the Seinhorst (1952) dehydration techniques. Dehydrated nematodes were mounted on clean non-greased slides with dehydrated glycerin as mountants. The specimens were studied, measurements taken and diagrams were drawn using a drawing tube attached to a microscope.

## RESULTS

## Systematics

## Aphelenchoides aeralis Chanu et al., 2015 (Figure 2, Table 1 & 3)

## Diagnosis

Female: Body contour S-shaped with fine cuticular striations having a lateral field with two incisures. The cephalic region set off with weak sclerotization, spear 15.3  $\mu$ m long with small basal thickenings. The median



Figure 1. Expanded map of Manipur.

oesophageal bulb is oblong with centered valvular apparatus. Oesophagous overlap intestine. Nerve ring behind the esophagus-intestinal junction. Excretory pore above nerve ring. Vagina with sphincter and raised vulval lips. Gonads monoprodelphic and oocytes are arranged in single rows. Spermatheca is filled with sperms and with uterine sac. Tail 42.5–48.2 µm long,conical, and with small single mucro.

Male: Body more curved at tail region. Testis long and outstretched. Spicule is typical of the genus. Dorsal limb without knob. The capitulum and rostrum very well developed. Post anal genital papillae, one pair situated above the tail terminus. Tail terminating into a long spine-like mucro. Remark: The morphometric details of the present species conform well with those described by Chanu et al. (2015).

## Aphelenchoides aligarheinsis Siddiqi et al., 1967 (Table 1)

## Diagnosis

Female: Body contour slightly curved. Cuticle fine, striations about 1.7  $\mu$ m at mid-body. Lip region set-off round. Lateral fields with four incisures. Stylet slender 10.2  $\mu$ m long with weakly developed basal knobs. Oesophagous typical of the genus. Excretory pore at level of the nerve ring. Ovary outstretched and oocytes arranged in single row. Spermatheca oblong with

CharacterA. aeralis Chanu et al., 2013A. aligar-hiensis Siddiqi et al., 1967		A. ba-guie Maslen, 1973 A. confusus Thorne & Malek, 1968		<i>A. dhanach-adi</i> Chanu et al., 2012	<i>A.longistyl-us</i> Chanu & Mohilal, 2014	
L	0.46-0.51	0.42-0.61	0.58-0.74	0.48-0.79	0.37–0.50	0.59–0.66
а	32.1-33.8	25.1–30.2	35.2–38.2	28.6-46.6	25.2–36.1	34.4–35.85
b	6.15-7.2	6.1–8.1	5.2-7.2	2.5-4.4	6.6–7.7	8.53–9.81
b´	3.66-4.1	-	3.6–4.2	-	3.3–7.4	6.03–6.49
с	9.8–10.1	11.4–16.1	14.2–16.2	23.8–38.8	5.3–7.4	13.67–14.58
c	5–6.4	2.5–3	3.8–4.4	2-2.4	6.6–9.2	5.6-6.14
V	66.7–68.2	64.2–71.8	62.8–68	76.8–78.6	57.9–62.9	67.78–69.76
G1	26.9–28.2	30.6–34.8	42.4–48.6	29–42.6	13.6–15.3	24.2
G <sub>2</sub>	-	-	14–16.2	4.3–18.2	-	39.39–43.08
Spear	15.3	10.2	10.2–12.8	10.2–17	-	-
Oesophago-us	125.8	73.1–81.6	146.4–152	134.7–241.4	95.2–129.0	98.61–102.07
Nerve ring	68			90.1–119	56.1–73.1	-
Excretory pore	62.9	52.7	82.4–94.2	86.7–115.6	61.2-85.0	-
Tail	42.5	35.7–39.1	38.4–46.2	8.5–10.2	62.9-85.0	41.52–48.44
ABD	8.5	11.9–15.3			8.5–10.2	6.92–8.65
PUS	-	35.7–37.1	92.4–96.4	20.4	-	-

Table 1. Morphometric data of female species of Aphelenchoides spp. from Manipur.

discoidal spermatozoa. The Uterine sac is five times body width in length. Tail 35.7–39.1  $\mu$ m long, elongate-conoid, ventrally arcuate, rounded with spine like mucro.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al., (2015).

## Aphelenchoides baguei Maslen, 1978 (Table 1)

#### Diagnosis

Female: Body contour slightly ventrally curved. Fine cuticularisation with four incisures throughout body length. Cephalic region set off flattened. Spear 10.2–12.8  $\mu$ m long with a small basal knob. Oesophagous typical of aphelenchoid. Mono-prodelphic reproductive system and ovary outstretched. Vulva is a transverse slit with slightly protruding lips. Oocytes in single row with a uterine sac. Tail 38.4–46.2  $\mu$ m long, about 4–5 times anal body width long, terminus with a small ventral mucro which is multi-papillate almost to its tips.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2015).

## Aphelenchoides confusus Thorne & Malek, 1968 (Table 1)

## Diagnosis

Female: Body contour gradually tapering near extremities. Cuticle with fine striations and lateral fields marked by four fine lines. Cephalic region set-off. Spear 10.2–17  $\mu$ m long. Oesophagous typical aphelenchoid with massive valvular apparatus. Nerve ring is behind the oesophageal bulb. Excretory pore at level of the nerve ring. Hemizonid posterior to excretory pore.

Vulva with protuberant labia and vagina directed forward. Uterus spheroid-shaped, filled with sperm. Ovary outstretched and post uterine sac collapsed. Tail 20.4–25.3  $\mu$ m long without a mucro.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2015).

## Aphelenchoides dhanachandi Chanu et al., 2012 (Figure 3, Table 1)

## Diagnosis

Female: Body contour slender ventrally curved. Lateral filed marked by three incisures. The body cuticle is fine. The cephalic framework is high. Spear 13.6–15.3  $\mu$ m long, slender with indistinct basal knobs. Oesophagous typical with tamarind seed-shaped median bulb. Nerve ring behind the median bulb, 59.5–69.0

Figure 2. Aphelenchoides aerialis Chanu et al., 2013: A—Anterior part of Female body | B—Entire body of Male | C—Entire body of Female | D—Anterior body part of male | E—Posterior end of body (enlarged) | F—Tail region of Female.

 $\mu m$  long. Excretory pore at the level of the nerve ring. Oesophageal gland lying dorsally along the intestine.

Monoprodelphic reproductive system and oocytes arranged in a single row with uterine sac. Vulva protrudes in some species. Tail 62.9–98.0  $\mu$ m long, highly curved ventrally tapering into a pointed terminus.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2012).

## Aphelenchoides longistylus Chanu & Mohilal, 2014 (Figure 4, Table 1 & 3)

## Diagnosis

Female: Body contour slightly curved with fine annulation. Lateral fields with four longitudinal lines merge into two lines at around the tail region. The cephalic region indistinctly set off with six equal lips. Spear 24.22  $\mu$ m long with indistinct basal knobs. Procorpus straight, median bulb spherical to pyriform in shape. Excretory pore at the base of the median bulb. The vulva is a transverse slit. The monoprodelphic reproductive system and oocytes are arranged in a single row. Uterine sac well developed. The tail gradually tapers into a cylindrical tube, terminating in a ventral prong tip.

Male: Slightly smaller than female. Tail slender with single terminal mucro. Spicules absent about 24.22  $\mu$ m long. Testis single, 335.62–342.45  $\mu$ m long.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2014).

## Aphelenchoides manipurensis Chanu & Mohilal, 2018 (Figure 5 & 6, Table 2 & 3)

## Diagnosis

Female: Body contour cylindrical, ventrally arcuate, the cephalic framework set off and flat. Cuticle marked by fine annulus. Lateral fields with two incisures. Spear short, 10.33–13.84  $\mu$ m long with small rounded basal knobs. The median bulb is spherical and basal bulb bifurcated. Excretory pore at 51.9–76.12  $\mu$ m from anterior end of body. Nerve ring behind median bulb,

Characters A. manipure-nsis Chanu & Mohilal, 2018 A. minor Seth & Sharma. 1986		A. neoechino- caudatus Chanu et al., 2012	A. neominoris Chanu & Mohilal, 2014	A. swarupi Seth & Sharma, 1986	A. vigor Thorne & Malek, 1968	
Length	0.294-0.461	0.28-0.36	0.53-0.60	0.35-0/43	0.52-0.68	0.44-0.49
а	24.28-38.14	18.2–26.2	25.5–28.6	36.29–40.6	34.4-42.4	24.8–29.3
b	6.3-8.65	4.8-7.2	8.3–9.0	4.23-6.51	5.8-8.2	2.7–4.8
b´	3.61-8.15	2.4-4.4	4.5-5.0	6.03–6.49	-	5.6-8.6
С	12.47-16.46	12.2–16.2	11.0–11.7	13.53–36.29	12.2–16.2	12.4–30.3
c′	3-4.25	2.2–2.6	5.3–5.4	1.75-3.75	2.2–2.6	1.5–3.5
V	68.88–71.69	66.4–72.2	64.4–64.9	69.95–71.65	66.4–72.2	69.2-88.6
Stylet	10.38–13.84	48.2–56.2	11.9	8.65	8.2-10.2	11.9–13.6
G <sub>1</sub>	24.34-33.01	4.2–5.2	-	41.87–50.0	32.4–36.4	33.5–45.9
G <sub>2</sub>	-	4.8-8.2	-	-	-	10.5–13.3
Oesophagous	-		119.0122.4	67.47–83.04	-	102–164.9
Nerve ring	-	-	66.3–68.0	-	-	74.8-88.4
Exc. Pore	-	47.6–54.6	62.9–64.6	-	-	61.2-78.2
Tail	20.76-31.14	22.4–30.8	45.9–54.4	12.11–25.95	32.3–37.4	15.3–35.7
ABD	6.92-8.65		8.5–10.2	6.92	8.5–12.4	10.2

## Table 2. Morphometric data of female species of Aphelenchoides spp. from Manipur.

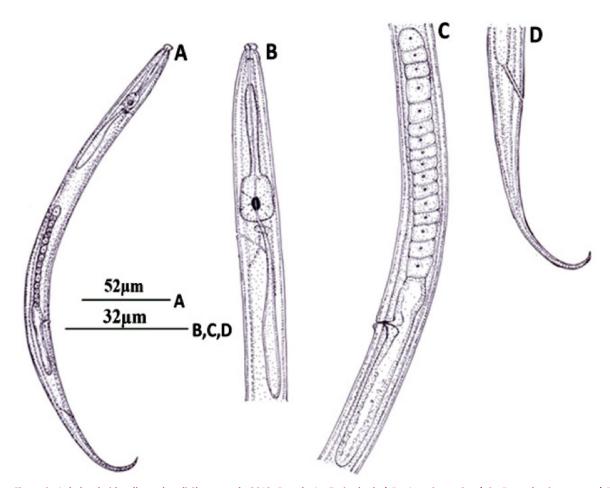


Figure 3. Aphelenchoides dhanachandi Chanu et al., 2012: Female A—Entire body | B—Anterior region | C—Reproductive system | D—Tail region.

ovary single, outstretched, and oocytes arranged in single row reaching the basal bulb. Spermatheca elongated oval with sperms. Uterine sac filled with sperms and ventral rounded tip. Vulva protuberant and vagina at right angle to the body. Tail curved ventrally with rounded tip with a small mucro at tip.

Male: Body ventrally curved. Tail conoid with mucronated lip. Testis 138.4–190.3  $\mu$ m. spermatocytes in single row, spicules simply arcuate, rostrum rounded, and prominent apex. Three pairs of sub ventral papillae present towards tip of spicule. Bursa and gubernaculum absent.

Remarks: The morphometric details of the present species conform well with those described by Chanu & Mohilal (2018).

Table 3. Morphometric data of male species of *Aphelenchoides* spp. from Manipur.

Characters	<i>A. aerialis</i> Chanu et al., 2013	A. longistylus Chanu & Mohilal, 2014	A. manipurensis Chanu & Mohilal, 2018
L	0.46–0.51	0.562-0.62	264.69-320.05
а	29–29.41	38.2–42.0	25.5–33.39
b	5.9–6.92	7.24–8.33	25.59–41.52
b´	3.4	3.25-5.60	3.47-4.02
с	8.7	13.54–17.45	12.0–13.91
c	4.2	4–6.2	2.4–6
т	46.71–47.8	92.61–102.84	50.89–61.09
Testis	46.71–47.8	335.62–342.45	138.4–190.3
Spicule	23.8–25.95	24.22	10.38–17.3
Tail	51.0–55.36	41.52-46.23	20.76–25.95
ABD	11.9–13.84	10.32	6.92-8.65
Stylet	15	24.22	10.38–13.84

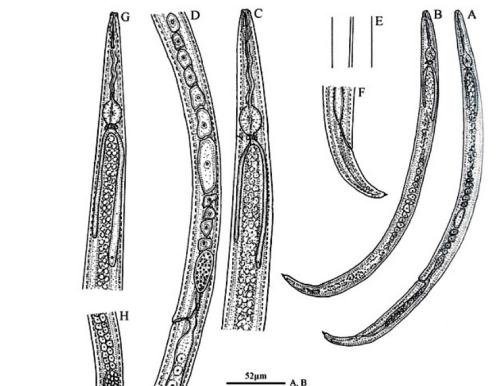


Figure 4. Aphelenchoides longistylus Chanu & Mohilal, 2014: Female. A—Entire body | C—Anterior body | D—Reproductive system | E— Lateral lines | F—Tail region. Male. | B—Entire body | G—Anterior body | H—Tail region.

C, D, E, F, G, H

32µm

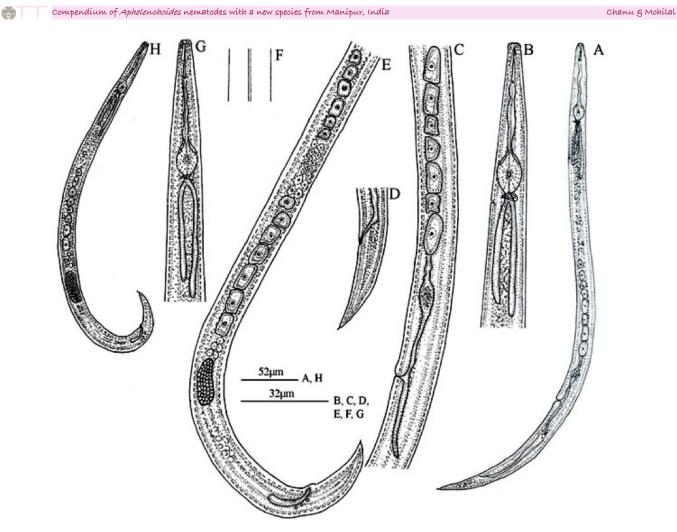


Figure 5. Aphelenchoides manipurensis Bina & Mohilal, 2018 from Manipur University Campus: Female. A—Entire body | C—Anterior body | D & I—Reproductive system (variation) | E—Body incisures | F—Anal region. Male. B—Male | Entire body | G—Anterior region | H—Posterior body.

Species	Host	Locality
A. aerialis Chanu et al., 2015	Pine tree, Pinus roxburghii Sarg	Nongpok Sekmai, Thounal District
A. aligarhiensis Siddiqi et al., 1967	Pine tree, Pinus kesiya Royle	Khuga Dam, Churchandpur District
A. baguei Maslen, 1973	Morus alba Lin, Pinus kesiya Royle	Kakwa Naorem Leikai, Imphal West Distrcit; Keibul Lamjao, Bishnupur District
A. confusus Thorne & Malek, 1968	<i>Morus alba</i> Linn	Matai garden, Imphal East District
A. minor Seth & Sharma, 1986	<i>Morus alba</i> Linn	Kalika Village, Irilbung, Kyamgei, Imphal East District; C. I. College, Bishnupur district; Govt. Silkfarm Wangbal , Thoubal district; Regional Tasar Research Station, Chingmeirong, Imphal West district.
A. swarupi Seth & Sharma, 1986	Morus alba Linn, Morus indica Linn, Pinus roxburghii Sarg	Regional Tasar Research Station, Chingmeirong, Imphal West District; Bishnupur ward no. 4, Bishnupur district; Nongpok Sekmai, Thoubal District
A. vigor Thorne & Malek, 1968	Morus indica Linn	Bishnupur ward no. 4, Bishnupur District
A. dhanachandi Chanu et al., 2012	Mulberry plant, Morus alba L.	Yurembam Rose Garden, Imphal West District
A. neoechinocaudatus Chanu et al., 2012	Mulberry plant, Morus alba L.	Yurembam Rose Garden, Imphal West District
A. neominoris Chanu & Mohilal, 2014	Orange plants	Sibilong, Chandel district
A. longistylus Chanu & Mohilal, 2014	Coconut tree (Cacos nucifera)	Ninghsing Khul, Jiri, Imphal West District
A. manipurensis Chanu & Mohilal, 2018	Rooten wood lock	Manipur University campus, Canchipur.

## Table 4. Species of Aphelenchoides spp. with their hosts and localities.

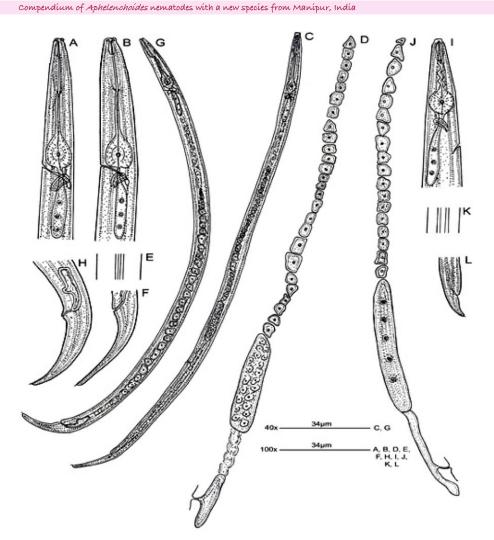


Figure 6. Aphelenchoides manipurensis Bina & Mohilal, 2018 from Nongpok Sekmai Pine Reserve Forest. Female: A—Entire body | B—Anterior body | C—Reproductive system | D—Anal region | F—Body incisures. Male: E—Reproductive system | G—Anterior region | H—Entire body.

## Aphelenchoides minor Seth & Sharma, 1986

(Table 2)

## Diagnosis

Female: Body contour straight to slightly curved ventrally. Lateral fields with three incisures. Cephalic framework set off without annulation. Spear 4.8–8.2  $\mu$ m long with indistinct basal thickenings. Oesophagous is aphelenchoid type with squarish muscular median bulb with a flat base and crescentric wave. Excretory pore at level of nerve ring. Vulva, a transverse slit with prominent lips. Oocytes are arranged in single row with a uterus. Tail 22.4–30.8  $\mu$ m long, rounded with ventral mucro.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2018).

## Aphelenchoides neoechinocaudatus Chanu et al., 2012 (Figure 7, Table 2)

## Diagnosis

Female: Body contour is slender with three lateral incisures. The cephalic region is slightly set off. Stylet slender 11.9  $\mu$ m long without basal swellings. Oesophagous typical, median bulb elongated pearshaped. Nerve ring 66.3–68.0  $\mu$ m long. Excretory pore at 62.9–64.4  $\mu$ m, at the level of the nerve ring. The oesophageal gland was dorsal to the intestine. Reproductive system monoprodelphic and oocytes arranged in a single row, and uterine sac well developed. Tail 45.9–54.4  $\mu$ m long, short, and pointed with a ventral mucro.

## Male: not found

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2012).

## Aphelenchoides neominoris Chanu & Mohilal, 2014 (Figure 8, Table 2)

## Diagnosis

Female: Body contour straight, tapering towards both extremities with four incisures in lateral field. Cephalic framework smooth and set-off. Spear 8.65  $\mu$ m long with distinct stylet knobs. Procorpus zig-zag, coiled, strongly rounded corpus with sclerotized plates and elongated gland lobe, dorsal to the intestine. The excretory pore is close to the nerve ring. Vulval lips protrude with an inclined vagina. Monoprodelphic reproductive system,

oocytes are arranged in a single row reaching up to the oesophageal bulb. Spermatheca large elongated and uterine sac empty. The anterior lip of the anus protrudes, the tail bluntly rounded, 12.11–25.95  $\mu$ m in length, with a small hair-like mucro.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu & Mohilal (2014).

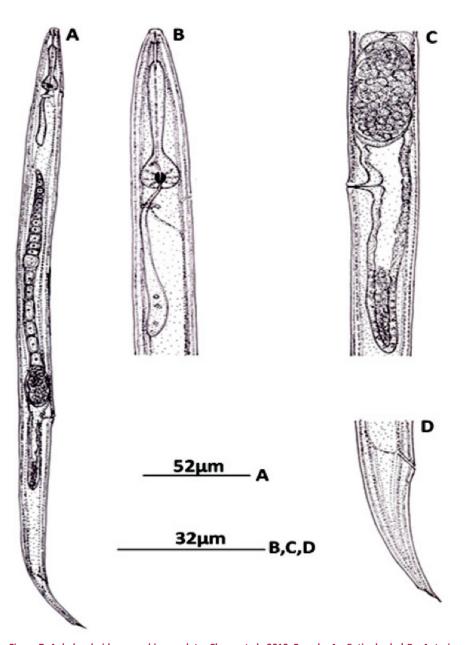


Figure 7. Aphelenchoides neoechinocaudatus Chanu et al., 2012. Female: A—Entire body | B—Anterior region | C—Reproductive system | D— Tail region. Photomicrographs: E—Anterior region | F—End bulb | G—Reproductive system | H—Tail region.

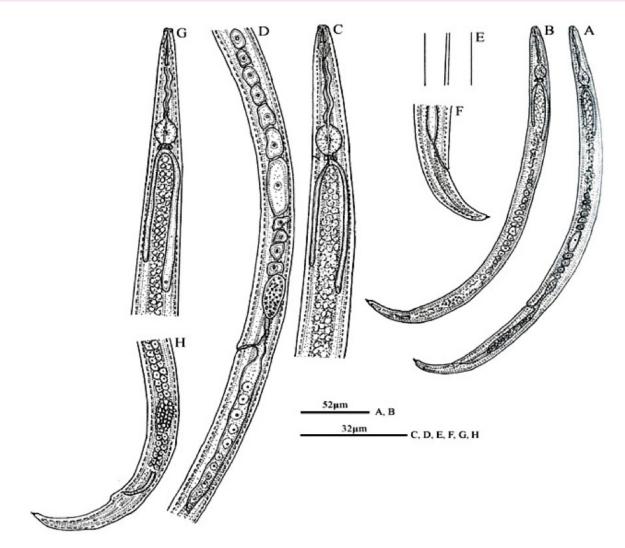


Figure 8. Aphelenchoides neominoris Chanu & Mohilal, 2014; A—Female whole body |B—Male whole body | C—Anterior portion of female | D—Body portion showing reproductive organs of female | E—Lateral lines | F—Female tail region | G—Anterior body region of male | H—Posterior body region of male showing testes.

# Aphelenchoides swarupi Seth & Sharma, 1986

Compendium of Aphelenchoides nematodes with a new species from Manipur, India

(Table 2)

## Diagnosis

Female: Body contour cylindrical to slightly curved, with three incisures in the lateral lines. Cephalic region set off without annules. Spear 8.2–10.2  $\mu$ m long with basal thickenings, procorpus muscular. Nerve ring at 72.8  $\mu$ m from anterior body. Excretory pore ventral at level of nerve ring. Tail 32.8–37.4  $\mu$ m long, bluntly rounded with a ventral mucro. Vulva a transverse slit with vulval lips. Post vulval uterine sac well developed.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2015).

## Aphelenchoides vigor Thorne & Malek,1968 (Table 2)

#### Diagnosis

Female: Body cylindrical with coarse annulations. Lateral filed with two incisures. The cephalic framework is set off by constriction. Spear 11.9–13.6  $\mu$ m long with distinct knobs. Nerve ring at 74.8–88.4  $\mu$ m long and excretory pore at 78.2  $\mu$ m from the anterior body region. Vulva sclerotized, overlapping with jointed flap. Ovary outstretched, tubular uterine sac filled with sperms. Tail 15.3–35.7  $\mu$ m long, arcuate, blunt tip without mucro.

Male: Not found.

Remarks: The morphometric details of the present species conform well with those described by Chanu et al. (2015).

The hosts and localities of all the species are

mentioned in Table 4.

Aphelenchoides oryzae sp. nov. (Figure 9, Table 5) urn:lsid:zoobank.org:act:223FC21F-A734-4794-B702-B396FA7F0D49

## Material examined

**Holotype**: Collected on August, 2015 from paddy fields (*Oryza sativa* L. growing field) by L. Bina chanu, from Thoubal Khekman, Thoubal District, Manipur with a longitude of 24.5036 and latitude of 93.9116. The specimen is deposited on nematode collection of Parasitology Section, Centre for Advanced Study in Life Sciences, Manipur University, Canchipur, Manipur under the Voucher no. ZoDMU\_MN02 with holotype female on slide FTY<sub>4</sub> 1.

**Paratype**: Females on slides  $FTY_4 \bigcirc 2-12$  and males on the slides  $FTY_4 \bigcirc 1-7$ , same data as holotype.

#### DESCRIPTIONS

Holotype female: Body straight, cylindrical, and robust upon fixation, 685.08  $\mu$ m long. Lip region offset with rounded sides & flattened anteriorly, 5.19  $\mu$ m wide and 1.7  $\mu$ m high, and smooth in appearance. Body elongate with fine transverse annulations, 0.8  $\mu$ m at mid body region. Lateral lines extend almost to tail tip with two ridges having four evenly spaced lines in the middle of the body.

Stylet is slender, 17.3  $\mu$ m long, the conus slightly shorter than the shaft with indistinct swellings. Median oesophageal bulb rounded to slightly oval with the refractive thickenings usually placed centrally, 13.84 $\mu$ m

Characters	Holotype female	Paratype females	Paratype males	
n	1	18	7	
Length	685.08	569.17-750.82 (675.045±58.74)	536.3-615.88 (576.09±39.79)	
а	44	36.55-44 (42.39±2.93)	34.44–39. 55 (36.99±2.65)	
b	7.33	6.18-7.33 (6.72±0.51)	5.74–5.83 (5.78±0.04)	
b´			-	
b <sub>1</sub>	9.65	9.65-11.12(10.26±0.55)	7.75–9.12 (8.43±0.68)	
с	36	16.45–36 (25.70± 8.59)	15.5–19.77 (17.63±2.13)	
C´	1.83	1.83-4 (3.144±0.94)	2.57-4 (3.28±0.71)	
Т	-	-	81.86–93.54 (94.24±0.69)	
V	69.44	68.20-70.7 (69.65±0.89)	-	
G <sub>1</sub>	49.24	29.95–49.24 (43.55±7.66)	-	
Post. Uterine sac	46.71	46.71-86.5 (62.97±15.61)	-	
PVS/ V-A%	24.54	24.54-51.02 (34.65±14.15)	-	
Oesophagous	93.42	91.69–114.18 (101.2±8.91)	91.69–114.18 (101.2±8.91)	
Stylet	17.3	17.3	17.3	
Lip width	5.19	5.19	5.19	
Lip height	1.73	1.73	1.73	
Median bulb length	13.84	10.38-13.84 (12.68±1.28)	10.38–13.84 (12.68±1.28)	
Median bulb diam.	8.65	6.92-8.65 (7.49± 0.81)	-	
Spicule	-	-	15.57–17.3(16.43±0.86)	
Ovary	337.35	224.9-337.35 (293.05±54.2)	-	
PUS/ VBD	3	3–6.25 (4.27±1.2)	-	
Nerve ring	74.39	69.2–76.12 (72.94±2.72)	69.2–76.12 (72.94±2.72)	
Excretory pore	72.66	72.66–77.85 (744.04±2.01)	72.66–77.85 (744.04±2.01)	
Spermatheca	29.41	29.41	-	
Rectum	8.65	5.19-8.65 (7.26± 1.69)	-	
Tail	19.03	19.03–41.52 (29.41± 8.59)	31.14–34.6 (32.87±1.73)	
ABD	10.38	8.65–10. 38 (9.51± 0.94)	8.65–12.11 (10.38± 1.73)	

Table 5. Morphometric data of species of Aphelenchoides oryzae sp. nov. All measurements in µm except L in mm.

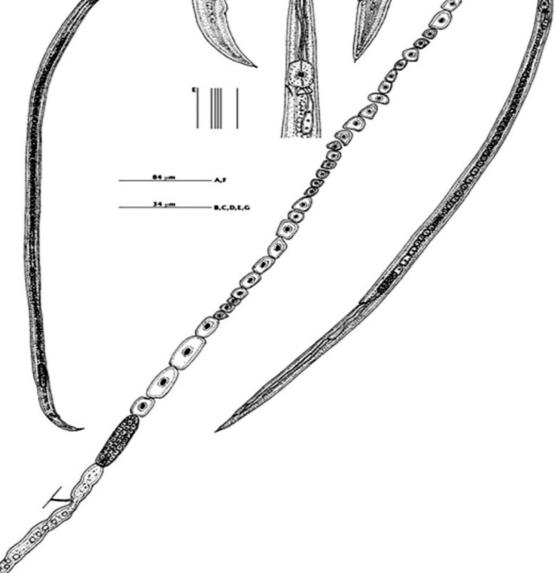


Figure 9. Aphelenchoides oryzae sp. nov.: Female, A—Entire body length | B—Anterior region | C—Female reproductive system | D—Tail region | E-Lateral lines | Male, F-Entire body length | G-Tail region.

high and 8.65µm across in length. Excretory pore and nerve ring, one to two body widths posterior to median bulb, excretory pore at 72.66µm from the anterior end and nerve ring at 74.39µm from the anterior end of the body. Oesophageal glands overlap the intestine.

Vulva is at about two thirds of the body length from the anterior end. Reproductive system with a single anterior ovary, oocytes arranged in single rows, with spermatheca, 29.41µm long filled with sperms and a prominent post-vulval uterine sac which usually extends just half the distance from the vulva to the anus. Post uterine sac is about 46.71µm in length.

Tail convex-conoid, 19.03µm long and straight, usually 10.35 µm anal body widths long with a rounded tail tip bearing a small terminal mucro.

Paratype males: Lip region, stylet and oesophagous similar to female. Tail ends curls ventrally through 45-90° when killed by heat and usually with simple terminal mucro. Spicules well developed; 15.57-17.3 (16.43 ± 0.86) µm long, the dorsal limb smoothly curved in its proximal half but flattened to concave tip; the ventral limb appears much weaker than the dorsal limb. The

#### Table 6. Characters differentiating Aphelenchoides oryzae sp. nov. from other related Aphelenchoides species.

Aphelenchoides spp.	Character differentiation				
A. blastophthorus Franklin, 1952	Shorter stylet (15–16 μm) with prominent knobs				
A. brassicae Edward & Misra, 1969	Excretory pore opposite median bulb base, female tail, shorter (c´=3) with longer mucro, spicules more smoothly curved.				
A. baguei Maslen, 1979	The female tail is ventrally concave with a longer mucro, and spicules more smoothly curved with a more prominent apex.				
A. hamatus Thorne & Malek, 1968 [After Vovlas, 1982]	Shorter stylet (12–13 $\mu$ m), female tail ventrally curved with a ventral mucro, spicule larger				
A. helophilus (de Man, 1880) Goodey, 1933	Female body length over 1mm, stylet 14–16 $\mu m$ with prominent knobs, spicules smoothly curved				
A. lanceolatus Tandon & Singh, 1974	Lip region continuous, stylet shorter (12.5–13 μm), female body thinner (a=33), shorter post–vulval sac, spicules smoothly curved				
A. lichenicola Siddiqi & Hawksworth, 1982	Female body was shorter (L=610 $\mu$ m), female tail longer (c'=3.5), spicules were characteristically swollen near the distal end of the dorsal limb				
A. lilium Yokoo, 1964	Excretory pore a body width posterior to nerve ring, shorter stylet (12.5 μm), shorter post–vulval sac, female tail ventrally curved, spicules smoothly curved				
A. saprophilus Franklin, 1957	Shorter female body (L= 546 $\mu m$ ), ventrally curved female tail and larger spicules				
A. sexlineatus Eroskenko ,1967	Shorter stylet (9 μm), shorter female body (L = 605–645 μm), longer post vulval sac, female tail with a longer mucro.				
A. submerses Truskova, 1973	Lip region narrower than the adjacent body; female tail more curved ventrally; excretory pore anterior to median bulb				
A. suipingensis Feng & Li, 1986	Female with the thinner body (a=32); excretory pore opposite median bulb base; female tail ventrally curved with hair-like mucro				
A. Tumulicaudatus Truskova, 1973	Lip region not offset, post vulval sac shorter; female tail with characteristic terminal swelling.				
A. nechaleos Hooper & Ibrahim, 1994	Female body longer (L=10.5–11.5 mm); shorter stylet (10.5–11.5 μm) and shorter tail (3.9–4.6 μm)				
A. paranechaleos Hooper & Ibrahim, 1994	Longer body (631–860 mm); shorter stylet (9.5–10.5μm); thinner body (a=37–46) and shorter tail (c´=2.6–3.6)				

rostrum and apex are moderately developed; a tangent drawn from the apex to the spicule tips is separated from the tangent from the apex through rostrum.

**Etymology**: The species name is derived from the host plant.

#### **Diagnosis and Relationships**

Aphelenchoides oryzae sp. nov. is characterized by narrow cylindrical body, adults being 569.17–750.82 (675.04±53.74)µm long with a stylet about 17.3 µm long with indistinct basal swellings, a prominent median bulb and short end bulb, 4 lateral lines throughout the body, and tail straight, convex-conoid with a simple terminal mucro.

Males are common and functional with prominal spicules with dorsal limb flattened to indent in its distal half and the tip curled ventrally. Lateral fields of adults usually with four lines.

In view of its association with paddy plants, *Aphelenchoides oryzae* sp. nov. might be confused with *Aphelenchoides besseyi* Christie, 1942 the rice nematode. However, *Aphelenchoides oryzae* sp. nov. is separated from *A. besseyi* Christie, 1942 in having a single, simple, tail mucro instead of three–four processes as in *A. besseyi*. The present species also has longer oesophagus and stylet than *A. besseyi* Christie, 1942 whereas oesophagus ranges from 64–68  $\mu$ m and stylet 10.0–12.5  $\mu$ m in *A. besseyi* Christie, 1942.

Aphelenchoides oryzae sp. nov. differed from other species of Aphelenchoides in having a female body length of 569.17–750.82 (675.04  $\pm$  58.74) µm, with a slender stylet length of 17.3 µm and a convex conoid tail with a simple terminal mucro with four lateral lines along the whole body length.

The differences of characters between closely related species of *Aphelenchoides* is provided in Table 6. Based on these morphometric differences the present species is reported as new to science.

The Shannon - Wiener species diversity index and Evenness for all the mentioned species are given in the table 7.

## CONCLUSIONS

The richness of the species in the region may be due to warm climatic conditions, suitable habitats, and hosts as well as due to the absence of drastic changes in the climatic conditions during the past few years. But the present work could not cover all the varied ecosystems of Manipur. Since, nematodes are soil dwelling living around the roots of plants as well as plant parasitic

Chann & Mohilal

(A)

Table 7. Shannon-Wiener species diversity index and evenness of the species of *Aphelenchoides*. Shannon-Wiener index is denoted by *H* and evenness by *E*. Total number of species is 255 and the Shannon-Wiener species index is 3.783.

Species	No. of individuals (n)	Proportion, pi=n/N	ln(pi)	pi ×ln(pi)	H = -Σ[(pi)×ln(pi)]	Ink	E=H / In(k)
A. aeralis Chanu et al., 2015	20	20/40=0.5	-0.6931	-0.346	0.346	3.688	0.093
A. aligarheinsis Siddiqi et al., 1967	5	5/35=0.1428	-1.9463	-0.277	0.277	3.555	0.077
A. baguei Maslen, 1973	18	18/90=0.2	-1.609	-0.321	0.321	4.499	0.071
A. confusus Thorne & Malek, 1968	10	10/32=-0.3125	-1.1631	-0.321	0.321	3.465	0.092
A. dhanachandi Chanu et al., 2012	12	12/53=0.2264	-1.485	-0.336	0.336	2.484	0.135
A. longistylus Chanu & Mohilal, 2014	17	17/46=0.3695	-0.995	-0.367	0.367	3.850	0.095
A. manipurensis Chanu & Mohilal, 2018	38	38/58=0.6551	0.422	-0.277	0.277	4.060	0.068
A. minor Seth & Sharma, 1986	51	51/200=0.255	-1.366	-0.348	0.348	5.298	0.065
A.neoechinocaudatus Chanu et al., 2012	4	4/53=0.0754	-2.5849	-0.194	0.194	3.970	0.048
A. neominoris Chanu & Mohilal, 2014	11	11/47=0.2340	-1.452	-0.339	0.339	3.850	0.088
A. swarupi Seth & Sharma, 1986	36	36/143=0.251	-1.382	-0.346	0.346	4.962	0.069
A. vigor Thorne & Malek, 1968	7	7/38=0.1842	-1.6917	-0.311	0.311	3.687	0.084
A. oryzae sp. nov.	26	26/43=0.4651	-0.7655	-0.356	0.356	5.366	0.074

## Key to the species of Aphelenchoides spp. from Manipur

1.	Cephalic region set-off from body
2.	Lateral fields with 2 incisures3Lateral fields with 3–4 incisures6
3.	Basal bulb elongated
4.	Vagina with lips A. aerialis Chanu et al., 2015 Vagina without lips
5.	Distinct spear knob, spear 11.9–13.6 μm long, arcuated blunt tail <i>A. vigor</i> Thorne & Malek, 1968 Indistinct spear knob, spear 17.3 μm long, tail convex-conoid with blunt tip <i>Aphelenchoides oryzae</i> sp. nov.
6.	Lateral fields with 3 incisures
7.	Tail tip pointed8Tail tip bluntly rounded
8.	Spear 4.8–8.2 μm, indistinct knob, tail 22.4–30.8 μm
9.	Tail with a mucro A. neoechinocaudatus Chanu et al., 2012 Tail without a mucro A. dhanachandi Chanu et al., 2012
10.	With simple single tail mucro       11         With multi-papillated tail mucro       A. baguei Maslen, 1978
11.	Uterine sac collapsed/absent A. confusus Thorne & Malek, 1968 Uterine sac well-developed
12.	Body contour curved, weak knob, tail elongated conoid with spine like mucro
	Body contour straight, tapering at extremities, knob distinct, tail blunt conoid with hair like mucro A. neominoris Chanu & Mohilal, 2014

forms, there is potential for availability of the organism in various other plant and tree varieties in different ecosystems of Manipur. There are various forest ecosystems found in Manipur with varied tree species along the Himalayan range. Further survey and in-depth taxonomic works incorporating molecular taxonomic techniques can reveal the rich diversity of the nematode group in Manipur.

#### REFERENCES

- Chanu, L.B., N. Mohilal & M. Manjur (2012). Two new species of Aphelenchoides (Nematoda: Aphelenchida: Aphelenchoidea: Aphelenchidae) from Manipur, India. Biologia 67(3): 530–534. https://doi.org/10.2478/s11756-012-0042-9
- Chanu, L.B. & N. Mohilal (2014). Two new Aphelenchoides Fischer, 1894 (Nematoda: Tylenchina) from Manipur, North East India. Biotropia 21(2): 93–99. https://doi.org/10.11598/btb.2014.21.2.3
- Chanu, L.B., N. Mohilal, L. Victoria & M. Manjur (2015). Eight known species of Aphelenchoides with description of a new species from Manipur, India. Journal of Parasitic Diseases 39: 225–233. https:// doi.org/10.1007/s12639-013-0323-4
- Chanu, L.B. & N. Mohilal (2018). Aphelenchoides manipurensis sp. nov. (Nematoda: Tylenchina) from Manipur, India. Journal of Parasitic Diseases 42(2): 291–296. https://doi.org/10.1007/S12639-018-0999-6
- **Christie, J.R. (1942).** A description of *Aphelenchoides besseyi* n. sp. the summer-dwarf nematode of strawberries with comments on the identity of *Aphelenchoides subtenuis* (Cobb 1926) and *Aphelenchoides hodsoni* Goodey 1936. *Proceedings of Helmintholological Society of Washington* 9: 82–84.
- **Cobb, N.A. (1918).** Estimating the nema population of soil with special reference to sugar-beet and root-gall nemas, *Heterodera radicicola* (Greef) Muller and with a description of *Tylencholaimus aequalis* n. sp. Agric Tech Circular no. 1, USA, Bureau of Plant Industry, U.S. Dept. of Agriculture, 48 pp.
- De Man, J.G. (1880). Die Eiheimischen, frei in der reinen Erde und im sliseen Wasser lebende Nematoden. Vorläufiger Bericht und descriptive - systematischer Theill. *Tijdschrift Der Nederlandsche Dicrkundige Vereeninging* 2: 78–196.
- Edward, J.C. & S.L. Misra (1969). Occurrence of some new species of Aphelenchoidea in the rhizosphere of certain field crops of Uttar Pradesh, India, with a note on an intersex. *Allahabad Farmer* 43: 1–6.

- Ferris, H., M. Eyre, R.C. Venette & S.S. Lau (1996). Population energetic of bacterial - feeding nematodes: stage-specific development and fecundity rates. *Soil Biology and Biochemistry* 28(3): 271–280. https://doi.org/10.1016/0038-0717(95)00127-1
- Ferris, H., S. Lau & R.C. Venette (1995). Population energetic of bacterial –feeding nematodes: Respiration and metabolic rates based on CO<sub>2</sub> production. *Soil Biology and Biochemistry* 27(3): 319– 330. https://doi.org/10.1016/0038-0717(94)00186-5
- Ferris, H. & R.C. Venette (1997). Thermal constrains to population growth of bacterial feeding nematodes. *Soil Biology and Biochemistry* 29(1): 63–74. https://doi.org/10.1016/0038–071(96)00259–3.
- Feng, Z-X. & L. Shao-Mei (1986). A new species of the genus Aphelenchoides (Tylenchida: Aphelenchoididae). Acta Zootaxon Sinica 11: 245–249.
- Goodey, T. (1933). Plant Parasitic Nematodes and The Diseases They Cause. E.P. Dutton & Co. Inc., New York, xx+306 pp.
- Hooper, D.J. & S.K. Ibrahim (1994). Aphelenchoides nechaleos n.sp. and A. paranechaleos n. sp. (Nematoda: Aphelenchoididae) from rice plants. Fundamentals of Applied Nematology 17(2): 153–160.
- Maslen, N.R. (1978). Six new nematode species from the maritime Antarctic. *Nematologica* 25: 288–308.
- Seinhorst, J.W. (1952). A rapid method for the transfer of nematodes from fixative to angydrous glycerin. *Nematologica* 4: 67–69.
- Seth, A. & N.K. Sharma (1986). Five new species of genus Aphelenchoides (Nematoda: Aphelenchida) infesting mushroom in Northern India. Journal of Soil Biology & Ecology 5: 14–19.
- Siddiqi, M.R. & D.L. Hawksworth (1982). Nematodes associated with galls on *Cladonia glauca*, including two new species. *Lichennologist* 14: 175–184.
- Siddiqi, M.R., S.I. Hussain & A.M. Khan (1967). Seinura propora n. sp. and Aphelenchoides aligarhiensis n. sp. (Nematoda: Tylenchida). Systematic Parasitologie 4: 257–262.
- Tandon, R.S. & S.P. Singh (1974). Nematode parasites of the common sponge gourd, Luffa cylinfrica from Lucknow. Geobios 1: 17–24.
- Thorne, G. & R.B. Malek (1968). Nematodes of the Northern Great Plains. Part I Tylenchida (Nemata: Secennentea). Agric Exp Statn, S Dakota St Univ, Brookings, S Dakota. Techn Bull 31: 111.
- Truskova, GM. (1973). Two new species of the genus Aphelenchoides s, (Nematoda: Aphelenchoididae). Zoological Journal 52: 596–598.
  Yeates, G.W. & T. Bongers (1999). Nematode diversity in agroecosystem, pp. 113–135. In: Paoletti (ed.). Invertebrate Biodiversity as Bioindicators of Sustainable Landscapes Practical Use of Invertebrates to assess sustainable land use. https://doi.org/10.1016/b978-0-444-50019-9.50010-8
- Yokoo, T. (1964). On a new species of *Aphelenchoides* (Aphelenchidae: Nematoda) a parasite of bulb of lily, from Japan. Agricultural Bulletien of Saga University, Japan 20: 67–69.



- Mr. Jatishwor Singh Irungbam, Biology Centre CAS, Branišovská, Czech Republic.
- Dr. Ian J. Kitching, Natural History Museum, Cromwell Road, Uk
- Dr. George Mathew, Kerala Forest Research Institute, Peechi, India Dr. John Noyes, Natural History Museum, London, UK
- Dr. Albert G. Orr, Griffith University, Nathan, Australia
- Dr. Sameer Padhye, Katholieke Universiteit Leuven, Belgium
- Dr. Nancy van der Poorten, Toronto, Canada
- Dr. Kareen Schnabel, NIWA, Wellington, New Zealand
- Dr. R.M. Sharma, (Retd.) Scientist, Zoological Survey of India, Pune, India
- Dr. Manju Siliwal, WILD, Coimbatore, Tamil Nadu, India
- Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India
- Dr. K.A. Subramanian, Zoological Survey of India, New Alipore, Kolkata, India Dr. P.M. Sureshan, Zoological Survey of India, Kozhikode, Kerala, India
- Dr. R. Varatharajan, Manipur University, Imphal, Manipur, India
- Dr. Eduard Vives, Museu de Ciències Naturals de Barcelona, Terrassa, Spain
- Dr. James Young, Hong Kong Lepidopterists' Society, Hong Kong
- Dr. R. Sundararaj, Institute of Wood Science & Technology, Bengaluru, India
- Dr. M. Nithyanandan, Environmental Department, La Ala Al Kuwait Real Estate. Co. K.S.C.,
- Kuwait Dr. Himender Bharti, Punjabi University, Punjab, India
- Mr. Purnendu Roy, London, UK
- Dr. Saito Motoki, The Butterfly Society of Japan, Tokyo, Japan
- Dr. Sanjay Sondhi, TITLI TRUST, Kalpavriksh, Dehradun, India
- Dr. Nguyen Thi Phuong Lien, Vietnam Academy of Science and Technology, Hanoi, Vietnam
- Dr. Nitin Kulkarni, Tropical Research Institute, Jabalpur, India
- Dr. Robin Wen Jiang Ngiam, National Parks Board, Singapore
- Dr. Lional Monod, Natural History Museum of Geneva, Genève, Switzerland.
- Dr. Asheesh Shivam, Nehru Gram Bharti University, Allahabad, India
- Dr. Rosana Moreira da Rocha, Universidade Federal do Paraná, Curitiba, Brasil Dr. Kurt R. Arnold, North Dakota State University, Saxony, Germany
- Dr. James M. Carpenter, American Museum of Natural History, New York, USA
- Dr. David M. Claborn, Missouri State University, Springfield, USA
- Dr. Kareen Schnabel, Marine Biologist, Wellington, New Zealand
- Dr. Amazonas Chagas Júnior, Universidade Federal de Mato Grosso, Cuiabá, Brasil
- Mr. Monsoon Jyoti Gogoi, Assam University, Silchar, Assam, India
- Dr. Heo Chong Chin, Universiti Teknologi MARA (UiTM), Selangor, Malaysia
- Dr. R.J. Shiel, University of Adelaide, SA 5005, Australia
- Dr. Siddharth Kulkarni, The George Washington University, Washington, USA
- Dr. Priyadarsanan Dharma Rajan, ATREE, Bengaluru, India
- Dr. Phil Alderslade, CSIRO Marine And Atmospheric Research, Hobart, Australia
- Dr. John E.N. Veron, Coral Reef Research, Townsville, Australia
- Dr. Daniel Whitmore, State Museum of Natural History Stuttgart, Rosenstein, Germany.
- Dr. Yu-Feng Hsu, National Taiwan Normal University, Taipei City, Taiwan
- Dr. Keith V. Wolfe, Antioch, California, USA Dr. Siddharth Kulkarni, The Hormiga Lab, The George Washington University, Washington,
- D.C., USA
- Dr. Tomas Ditrich, Faculty of Education, University of South Bohemia in Ceske Budejovice, Czech Republic
- Dr. Mihaly Foldvari, Natural History Museum, University of Oslo, Norway
- Dr. V.P. Uniyal, Wildlife Institute of India, Dehradun, Uttarakhand 248001, India
- Dr. John T.D. Caleb, Zoological Survey of India, Kolkata, West Bengal, India
- Dr. Priyadarsanan Dharma Rajan, Ashoka Trust for Research in Ecology and the Environment (ATREE), Royal Enclave, Bangalore, Karnataka, India

#### Fishes

- Dr. Neelesh Dahanukar, IISER, Pune, Maharashtra, India
- Dr. Topiltzin Contreras MacBeath, Universidad Autónoma del estado de Morelos, México
- Dr. Heok Hee Ng, National University of Singapore, Science Drive, Singapore
- Dr. Rajeev Raghavan, St. Albert's College, Kochi, Kerala, India
- Dr. Robert D. Sluka, Chiltern Gateway Project, A Rocha UK, Southall, Middlesex, UK
- Dr. E. Vivekanandan, Central Marine Fisheries Research Institute, Chennai, India
- Dr. Davor Zanella, University of Zagreb, Zagreb, Croatia
- Dr. A. Biju Kumar, University of Kerala, Thiruvananthapuram, Kerala, India Dr. Akhilesh K.V., ICAR-Central Marine Fisheries Research Institute, Mumbai Research
- Centre, Mumbai, Maharashtra, India Dr. J.A. Johnson, Wildlife Institute of India, Dehradun, Uttarakhand, India
- Dr. R. Ravinesh, Gujarat Institute of Desert Ecology, Gujarat, India
- Amphibians

Dr. Sushil K. Dutta, Indian Institute of Science, Bengaluru, Karnataka, India Dr. Annemarie Ohler, Muséum national d'Histoire naturelle, Paris, France

#### Reptiles

- Dr. Gernot Vogel, Heidelberg, Germany
- Dr. Raju Vyas, Vadodara, Gujarat, India
- Dr. Pritpal S. Soorae, Environment Agency, Abu Dubai, UAE.
- Prof. Dr. Wayne J. Fuller, Near East University, Mersin, Turkey Prof. Chandrashekher U. Rivonker, Goa University, Taleigao Plateau, Goa. India
- Dr. S.R. Ganesh, Chennai Snake Park, Chennai, Tamil Nadu, India

Dr. Himansu Sekhar Das, Terrestrial & Marine Biodiversity, Abu Dhabi, UAE

------

Journal of Threatened Taxa is indexed/abstracted in Bibliography of Systematic Mycology, Biological Abstracts, BIOSIS Previews, CAB Abstracts, EBSCO, Google Scholar, Index Copernicus, Index Fungorum, JournalSeek, National Academy of Agricultural Sciences, NewJour, OCLC WorldCat, SCOPUS, Stanford University Libraries, Virtual Library of Biology, Zoological Records.

#### NAAS rating (India) 5.64

- Birds
- Dr. Hem Sagar Baral, Charles Sturt University, NSW Australia Mr. H. Byju, Coimbatore, Tamil Nadu, India Dr. Chris Bowden, Royal Society for the Protection of Birds, Sandy, UK
- Dr. Priya Davidar, Pondicherry University, Kalapet, Puducherry, India Dr. J.W. Duckworth, IUCN SSC, Bath, UK
- Dr. Rajah Jayapal, SACON, Coimbatore, Tamil Nadu, India
- Dr. Rajiv S. Kalsi, M.L.N. College, Yamuna Nagar, Haryana, India
- Dr. V. Santharam, Rishi Valley Education Centre, Chittoor Dt., Andhra Pradesh, India
- Dr. S. Balachandran, Bombay Natural History Society, Mumbai, India
- Mr. J. Praveen, Bengaluru, India
- Dr. C. Srinivasulu, Osmania University, Hyderabad, India
- Dr. K.S. Gopi Sundar, International Crane Foundation, Baraboo, USA
- Dr. Gombobaatar Sundev, Professor of Ornithology, Ulaanbaatar, Mongolia
- Prof. Reuven Yosef, International Birding & Research Centre, Eilat, Israel Dr. Taej Mundkur, Wetlands International, Wageningen, The Netherlands
- Dr. Carol Inskipp, Bishop Auckland Co., Durham, UK
- Dr. Tim Inskipp, Bishop Auckland Co., Durham, UK Dr. V. Gokula, National College, Tiruchirappalli, Tamil Nadu, India
- Dr. Arkady Lelej, Russian Academy of Sciences, Vladivostok, Russia
- Dr. Simon Dowell, Science Director, Chester Zoo, UK
- Dr. Mário Gabriel Santiago dos Santos, Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, Vila Real, Portugal
- Dr. Grant Connette, Smithsonian Institution, Royal, VA, USA
- Dr. P.A. Azeez, Coimbatore, Tamil Nadu, India

#### Mammals

- Dr. Giovanni Amori, CNR Institute of Ecosystem Studies, Rome, Italy
- Dr. Anwaruddin Chowdhury, Guwahati, India
- Dr. David Mallon, Zoological Society of London, UK
- Dr. Shomita Mukherjee, SACON, Coimbatore, Tamil Nadu, India

Dr. Nishith Dharaiya, HNG University, Patan, Gujarat, India

Dr. Dan Challender, University of Kent, Canterbury, UK

- Dr. Angie Appel, Wild Cat Network, Germany
- Dr. P.O. Nameer, Kerala Agricultural University, Thrissur, Kerala, India
- Dr. Ian Redmond, UNEP Convention on Migratory Species, Lansdown, UK
- Dr. Heidi S. Riddle, Riddle's Elephant and Wildlife Sanctuary, Arkansas, USA

Dr. Honnavalli N. Kumara, SACON, Anaikatty P.O., Coimbatore, Tamil Nadu, India

Dr. Justus Joshua, Green Future Foundation, Tiruchirapalli, Tamil Nadu, India

Dr. Jim Sanderson, Small Wild Cat Conservation Foundation, Hartford, USA

Dr. David Mallon, Manchester Metropolitan University, Derbyshire, UK

Dr. Brian L. Cypher, California State University-Stanislaus, Bakersfield, CA

Dr. Hemanta Kafley, Wildlife Sciences, Tarleton State University, Texas, USA

Dr. Mandar S. Paingankar, University of Pune, Pune, Maharashtra, India (Molecular)

Dr. Jack Tordoff, Critical Ecosystem Partnership Fund, Arlington, USA (Communities)

Dr. Rayanna Hellem Santos Bezerra, Universidade Federal de Sergipe, São Cristóvão, Brazil Dr. Jamie R. Wood, Landcare Research, Canterbury, New Zealand Dr. Wendy Collinson-Jonker, Endangered Wildlife Trust, Gauteng, South Africa

Dr. L.D. Singla, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, India

Dr. S.S. Talmale, Zoological Survey of India, Pune, Maharashtra, India Prof. Karan Bahadur Shah, Budhanilakantha Municipality, Kathmandu, Nepal Dr. Susan Cheyne, Borneo Nature Foundation International, Palangkaraja, Indonesia

Dr. Aniruddha Belsare, Columbia MO 65203, USA (Veterinary)

Dr. Ulrike Streicher, University of Oregon, Eugene, USA (Veterinary)

Dr. Hari Balasubramanian, EcoAdvisors, Nova Scotia, Canada (Communities)

Dr. Rajeshkumar G. Jani, Anand Agricultural University, Anand, Gujarat, India Dr. O.N. Tiwari, Senior Scientist, ICAR-Indian Agricultural Research Institute (IARI), New

Dr. Rupika S. Rajakaruna, University of Peradeniya, Peradeniya, Sri Lanka Dr. Bahar Baviskar, Wild-CER, Nagpur, Maharashtra 440013, India

Due to pausity of space, the list of reviewers for 2018-2020 is available online.

The opinions expressed by the authors do not reflect the views of the

boundaries shown in the maps by the authors.

Print copies of the Journal are available at cost. Write to:

c/o Wildlife Information Liaison Development Society,

43/2 Varadarajulu Nagar, 5th Street West, Ganapathy, Coimbatore,

Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political

Dr. H. Raghuram, The American College, Madurai, Tamil Nadu, India

Dr. Spartaco Gippoliti, Socio Onorario Società Italiana per la Storia della Fauna "Giuseppe

- Dr. Karin Schwartz, George Mason University, Fairfax, Virginia.
- Dr. Lala A.K. Singh, Bhubaneswar, Orissa, India
- Dr. Mewa Singh, Mysore University, Mysore, India Dr. Paul Racey, University of Exeter, Devon, UK

Dr. Paul Bates, Harison Institute, Kent, UK

Altobello", Rome, Italy

**Other Disciplines** 

Delhi, India

Reviewers 2020-2022

The Managing Editor, JoTT,

Tamil Nadu 641006, India ravi@threatenedtaxa.org





The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

## ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

## October 2023 | Vol. 15 | No. 10 | Pages: 23931-24150 Date of Publication: 26 October 2023 (Online & Print) DOI: 10.11609/jott.2023.15.10.23931-24150

www.threatenedtaxa.org

#### Articles

Echolocation call characterization of insectivorous bats from caves and karst areas in southern Luzon Island, Philippines

– Renz Angelo Duco, Anna Pauline de Guia, Judeline Dimalibot, Phillip Alviola & Juan Carlos Gonzalez, Pp. 23931-23951

Seasonality, diversity, and forest type associations of macro moths (Insecta: Lepidoptera: Heterocera) in the Shiwalik landscape of northern India and its conservation implications

– Arun Pratap Singh & Lekhendra, Pp. 23952–23976

#### Vertebrate assemblages on fruiting figs in the Indian eastern Himalaya's Pakke Wildlife Sanctuary

- Akangkshya Priya Gogoi, Janmejay Sethy, Awadhesh Kumar, Dipika Parbo, Murali Krishna Chatakonda & Ajay Maletha, Pp. 23977–23989

#### Communications

From the Arabian Peninsula to Indian shores: Crab Plover Dromas ardeola Paykull, 1805 (Aves: Charadriiformes: Dromadidae) breeding at Point Calimere, India

- H. Byju, N. Raveendran & K.M. Aarif, Pp. 23990-23995

Assessing avian diversity and conservation status in Dighal Wetlands, Haryana, India

– Parul & Parmesh Kumar, Pp. 23996–24008

Studies on the response of House Sparrow Passer domesticus to artificial nestboxes in rural Arakkonam and Nemili taluks, Vellore District, Tamil Nadu, India – M. Pandian, Pp. 24009–24015

### Threat assessment and conservation challenges for the herpetofaunal diversity of Dampa Tiger Reserve, Mizoram, India

- Sushanto Gouda, Ht. Decemson, Zoramkhuma, Fanai Malsawmdawngliana, Lal Biakzuala & Hmar Tlawmte Lalremsanga, Pp. 24016–24031

Taxonomy and conservation status of swamp eels (Synbranchiformes: Synbranchidae) of West Bengal, India - Ram Krishna Das, Pp. 24032-24042

Sacred river of Pune: boon or bane for the diversity of aquatic beetles (Insecta: Coleoptera)

– Rita Deb, Pallavi Takawane & K.A Subramanian, Pp. 24043–24053

#### Fine structure of sensilla on the proboscis of the Indian Honey Bee Apis cerana indica Fabricius (Insecta: Hymenoptera: Apidae)

– A.G. Suhas Krishna, Shamprasad Varija Raghu & Rajashekhar K. Patil, Pp. 24054-24062

## A compendium of Aphelenchoides (Fischer, 1894) (Nematoda: Tylenchina: Aphelenchoidea) nematodes with the description of a new species from Manipur, India

– Loukrakpam Bina Chanu & Naorem Mohilal, Pp. 24063–24078

Efficacy of levamisole and oxyclozanide treatment on gastrointestinal nematodes of ungulates at the Central Zoo, Nepal

- Pratik Kiju, Amir Sadaula, Parbat Jung Thapa & Chiranjibi Prasad Pokheral, Pp. 24079-24085

Ocimum gratissimum L. ssp. gratissimum var. macrophyllum Brig. (Lamiaceae: Nepetoideae: Ocimeae) a new record from northeastern India - Mamita Kalita, Nilakshee Devi & Diganta Narzary, Pp. 24086-24091

The study of biogeographic patterns of the genus Parmotrema in Wayanad District, Kerala with a new record in India

- Bibin Joseph, Edathum Thazhekuni Sinisha, Valiya Thodiyil Jaseela, Harshid Pulparambil & Nediyaparambu Sukumaran Pradeep, Pp. 24092-24103

#### Review

Diversity of Calliphoridae and Polleniidae (Diptera) in the Himalaya, India - Meenakshi Bharti, Pp. 24104-24115

#### Short Communications

First photographic evidence of mange manifestation in Panna Tiger Reserve, India

- Supratim Dutta & Krishnamurthy Ramesh, Pp. 24116-24119

New locality record of Forest Spotted Gecko Cyrtodactylus (Geckoella) cf. speciosus (Beddome, 1870) (Reptilia: Squamata: Gekkonidae) from Thanjavur, in the eastern coastal plains of Tamil Nadu, India – Gopal Murali, Pp. 24120–24124

Preliminary observations of moth (Lepidoptera) fauna of Purna Wildlife Sanctuary, Guiarat, India Preeti Choudhary & Indu Sharma, Pp. 24125–24130

On the occurrence of Audouinella chalybea (Roth) Bory, 1823, a rare freshwater red algae (Florideophyceae: Acrochaetiales: Audouinellaceae) from eastern Himalaya, India

- Jai Prakash Keshri & Jay Mal, Pp. 24131-24134

Addition of four invasive alien plant species to state flora of Mizoram, India - Lal Tlanhlui, Margaret Lalhlupuii, Sanatombi Devi Yumkham & Sandhyarani Devi Khomdram, Pp. 24135-24139

#### Notes

First sighting record of Western Reef-Heron Egretta gularis (Bosc, 1792) (Aves: Pelecaniformes: Ardeidae) from Jammu & Kashmir. India

- Parvaiz Yousuf, Semran Parvaiz, Nisheet Zehbi, Sabia Altaf, Showkat Maqbool, & Mudasir Mehmood Malik, Pp. 24140–24143

Rare desmid genus Bourrellyodesmus Compère (Chlorophyceae: Desmidiales: Desmidiaceae) in India with description of a new species (Bourrellyodesmus indicus Das & Keshri sp. nov.) from eastern Himalaya, India - Debjyoti Das & Jai Prakash Keshri, Pp. 24144-24147

Threats faced by Humboldtia bourdillonii Prain (Magnoliopsida: Fabales: Fabaceae), an endangered tree endemic to the southern Western Ghats, India - Jithu K. Jose & K. Anuraj, Pp. 24148-24150



